



## Testing Tips

- ✚ Be precise and consistent with each step to limit variability in technique between tests.
- ✚ If the soil or compost is very wet or clay-like consistency, break up any large chunks with the spatula included with your kit. Then, using both sifters, fit one inside the other, and “grind” the soil to remove any debris. If this does not work, simply remove any debris or large particles by hand and proceed with testing.
- ✚ In the pipetting stage of the test, draw up soil solution 1 inch from the surface.
- ✚ Avoid drawing up any floating debris or particles into the pipette.
- ✚ Avoid the sides of the tube when taking up the solution with the pipette.
- ✚ Make sure you’re pipetting at a consistent angle to limit variability in drop volume size.
- ✚ Place the test card on the enclosed coaster before scanning the test card with your smartphone.
- ✚ Perform test in similar lighting conditions to limit any outside variables.

## Sampling Tips

- ✚ Limit variability in sampling conditions. We recommend taking samples around the same time of day and in similar weather conditions and moisture levels – unless doing a specific study to compare conditions.
- ✚ Sample in or as close to the plant-soil microbiome as possible – this can include the soil attached to the roots (the rhizosphere). If sampling the rhizosphere, be very careful not to damage the roots and plant.
- ✚ Collect field moist soil samples at a depth between 2-5 inches depending on root length, being sure to take samples at consistent depths.
- ✚ Test the sample immediately after collecting. If you’re unable to test the sample the same day of collection, place sample in a sealed plastic bag for up to 72 hours. If more time is needed, you can refrigerate the sealed samples for up to 4 weeks. Make sure storing conditions are the same for each sample before testing to minimize any variation.